

# **Human Angiotensin Converting Enzyme 2 (ACE 2) ELISA**

**Product Data Sheet** 

Cat. No.: RAG006R

For Research Use Only

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- This kit is manufactured by:
  BioVendor Laboratorní medicína a.s.
- Use only the current version of Product Data Sheet enclosed with the kit!

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#### 1. INTENDED USE

ACE2 (human) ELISA Kit is to be used for the *in vitro* quantitative determination of human ACE2 in urine and cell culture supernatant. This ELISA Kit is for research use only.

### 2. HANDLING, STORAGE

- Reagent must be stored at 2-8°C when not in use.
- Plate and reagents should be at room temperature before use.
- Do not expose reagents to temperatures greater than 25°C.

#### 3. INTRODUCTION

By EST database searching for sequences showing homology to the zinc metalloprotease angiotensin-I converting enzyme, a full-length ACE2 cDNA, originally called ACEH, was isolated, which encoded a deduced 805-amino acid protein that shares approximately 40% identity with the N- and C-terminal domains of ACE. ACE2 contains a potential 17-amino acid N-terminal signal peptide and a putative 22-amino acid C-terminal membrane anchor. Northern blot analysis detected high expression of ACE2 in kidney, testis, and heart, and moderate expression in colon, small intestine, and ovary. Tipnis et al. (1) expressed a soluble, truncated form of ACE2 lacking transmembrane and cytosolic domains in CHO cells and found that it produced a glycosylated protein that was able to cleave angiotensin I and angiotensin II, but not bradykinin. Boehm and Nabel (2) showed whereas ACE converts angiotensin I to angiotensin II, which has 8 amino acids, ACE2 converts angiotensin I to angiotensin 1-9, which has 9 amino acids. This can then further be converted by ACE to a shorter peptide, angiotensin 1-7, which is a blood vessel dilator. Using ACE2 null mice, Crackower et al. showed that ACE2 was critically involved in a cardiac contractility (3). Li et al. (4) identified ACE2, isolated from SARS coronavirus-permissive Vero E6 cells, that efficiently binds the S1 domain of the SARS coronavirus S protein. It was shown that when ACE2 was engaged with S protein ACE2 surface expression was downregulated (5), increasing lung levels of angiotensin II, which was proposed to give rise to a severe lung injury. Since ACE2 is detected in urine (1), measurement of shedded ACE2 may provide a novel clue to renal & cardiovascular function of ACE2.

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#### 4. TEST PRINCIPLE

This assay is a sandwich Enzyme Linked-Immunosorbent Assay (ELISA) for quantitative determination of human ACE2 in biological fluids. A polyclonal antibody specific for ACE2 has been precoated onto the 96-well microtiter plate. Standards and samples are pipetted into the wells for binding to the coated antibody. After extensive washing to remove unbound compounds, ACE2 is recognized by the addition of a biotinylated polyclonal antibody specific for ACE2 (Detection Antibody). After removal of excess biotinylated antibody, HRP labeled streptavidin (Detector) is added. Following a final washing, peroxidase activity is quantified using the substrate 3,3',5,5'-tetramethylbenzidine (TMB). The intensity of the color reaction is measured at 450 nm after acidification and is directly proportional to the concentration of ACE2 in the samples.

#### 5. TECHNICAL HINTS

- It is recommended that all standards, QC sample and samples be run in duplicate.
- Do not combine leftover reagents with those reserved for additional wells.
- Reagents from the kit with a volume less than 100 µl should be centrifuged.
- Residual wash liquid should be drained from the wells after last wash by tapping the plate on absorbent paper.
- Crystals could appear in the 10X solution due to high salt concentration in the stock solutions. Crystals are readily dissolved at room temperature or at 37°C before dilution of the buffer solutions.
- Once reagents have been added to the 8-well strips, DO NOT let the strips DRY at any time during the assay.
- Keep TMB Substrate Solution protected from light.
- The Stop Solution consists of phosphoric acid. Although diluted, the Stop Solution should be handled with gloves, eye protection and protective clothing.

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# 6. REAGENT SUPPLIED

Kit Components	Quantity
1 plate coated with human ACE2 Antibody	12 x 8-well strips
1 bottle Wash Buffer 10X	50 ml
1 bottle Diluent 5X	50 ml
1 bottle Detection Antibody	12 ml
1 vial Detector 100X (HRP Labeled Streptavidin)	150 µl
1 vial human ACE2 Standard (lyophilized)	50 ng
1 vial human ACE2 QC sample (lyophilized)	
1 bottle TMB Substrate Solution	12 ml
1 bottle Substrate Solution II (Peroxidase)	6 ml
1 bottle Stop Solution	12 ml
3 plate sealers (plastic film)	

# 7. MATERIALS REQUIRED BUT NOT SUPPLIED

- Microtiterplate reader at 450 nm, with the correction wavelength set at 540 nm or 570 nm
- Calibrated precision single and multi-channel pipettes. Disposable pipette tips
- Deionized water
- Microtubes or equivalent for preparing dilutions
- Disposable plastic containers for preparing working buffers
- Plate washer: automated or manual
- Glass or plastic tubes for diluting and aliquoting standard

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#### 8. PREPARATION OF REAGENTS

**NOTE:** Prepare just the appropriate amount of the buffers necessary for the assay.

- Wash Buffer 10X has to be diluted with deionized water 1:10 before use (e.g. 50 ml Wash Buffer 10X + 450 ml water) to obtain Wash Buffer 1X.
- <u>Diluent 5X</u> has to be diluted with deionized water 1:5 before use (e.g. 50 ml Diluent 5X + 200 ml water) to obtain Diluent 1X.
- <u>Detector 100X (HRP Labeled Streptavidin)</u> has to be diluted to the working concentration by adding 120 µl in 12 ml of Diluent 1X (1:100).

**NOTE:** The diluted Detector is used within one hour of preparation.

- Human ACE2 Standard (STD) has to be reconstituted with 1 ml of deionized water.
  - o This reconstitution produces a stock solution of 50 ng/ml. Mix the standard to ensure complete reconstitution and allow the standard to sit for a minimum of 15 minutes. Mix well prior to making dilutions.

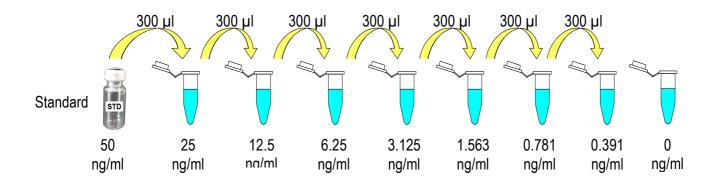
**NOTE:** The reconstituted standard is aliquoted and stored at -20°C

- o Dilute the standard protein concentrate (STD) (**50 ng/ml**) in Diluent 1X. A seven-point standard curve using 2-fold serial dilutions in Diluent 1X is recommended.
- Suggested standard points are:
- 25, 12.5, 6.25, 3.125, 1.563, 0.781, 0.391 and 0 ng/ml.
- Human ACE2 QC sample has to be reconstituted with 1 ml of deionized water.
  - Refer to the Certificate of Analysis for current QC sample concentration. Mix the QC sample to ensure complete reconstitution and allow the QC sample to sit for a minimum of 15 minutes. The reconstituted QC sample is ready to use, do not dilute it.

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#### Dilute further for the standard curve:

To obtain	Add	Into		
25 ng/ml	300 µl of ACE2 (50 ng/ml)	300 µl of Diluent 1X		
<b>12.5 ng/ml</b> 300 μl of ACE2 (25 ng/ml)		300 μl of Diluent 1X		
<b>6.25 ng/ml</b> 300 μl of ACE2 (12.5 ng/ml)		300 µl of Diluent 1X		
3.125 ng/ml	300 µl of ACE2 (6.25 ng/ml)	300 µl of Diluent 1X		
<b>1.563 ng/ml</b> 300 μl of ACE2 (3.125 ng/ml)		300 µl of Diluent 1X		
<b>0.781 ng/ml</b> 300 μl of ACE2 (1.563 ng/ml)		300 µl of Diluent 1X		
0.391 ng/ml	300 µl of ACE2 (0.781 ng/ml)	300 µl of Diluent 1X		
0 ng/ml 300 μl of Diluent 1X		Empty tube		



# 9. PREPARATION OF SAMPLES

**Urine:** Aseptically collect the urine of the day, voided directly into a sterile container. Assay immediately or aliquot and store at  $\leq$  -20°C. Avoid repeated freeze/thaw cycles.

**Urine or Cell Culture Supernatant** have to be diluted in Diluent 1X. Samples containing visible precipitates must be clarified before use.

**NOTE:** As a starting point, 1/2 dilution of urine is recommended! If samples fall the outside range of assay, a lower or higher dilution may be required!

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#### 10. ASSAY PROCEDURE

- 1. .Determine the number of 8-well strips needed for the assay and insert them in the frame for current use. The extra strips should be resealed in the foil pouch bag and stored at 4°C.
  - **NOTE:** Remaining 8-well strips coated with ACE2 antibody when opened can be stored at 4°C for up to 1 month.
- 2. Add 100 µl of the different standards into the appropriate wells in duplicate! At the same time, add 100 µl of diluted urine or cell culture supernatant samples in duplicate to the wells (see 8.1. Preparation and Storage of Reagents and 8.2. Preparation of Samples).
- 3. Cover the plate with plate sealer and incubate for 1 hour at 37°C.
- 4. Aspirate the coated wells and add 300 µl of Wash Buffer 1X using a multichannel pipette or auto-washer. Repeat the process for a total of three washes. After the last wash, complete removal of liquid is essential for good performance.
- 5. Add 100 µl to each well of the Detection Antibody.
- 6. Cover the plate with plate sealer and incubate for 1 hour at 37°C.
- 7. Aspirate the coated wells and add 300 µl of Wash Buffer 1X using a multichannel pipette or auto-washer. Repeat the process for a total of three washes. After the last wash, complete removal of liquid is essential for good performance.
- 8. Add 100 µl to each well of the diluted Detector (see 8.1. Preparation and Storage of Reagents).
- 9. Cover the plate with plate sealer and incubate for **1 hour at 37°C**.
- 10. Aspirate the coated wells and add 300 µl of Wash Buffer 1X using a multichannel pipette or auto-washer. Repeat the process for a total of five washes. After the last wash, complete removal of liquid is essential for good performance.
- 11. Add 100 µl to each well of TMB Substrate Solution.
- 12. Allow the color reaction to develop at room temperature (RT°C) in the dark for 30 minutes.
- 13. Stop the reaction by adding 100 µl of Stop Solution. Tap the plate gently to ensure thorough mixing. The substrate reaction yields a blue solution that turns yellow when Stop Solution is added.

#### ! CAUTION: CORROSIVE SOLUTION!

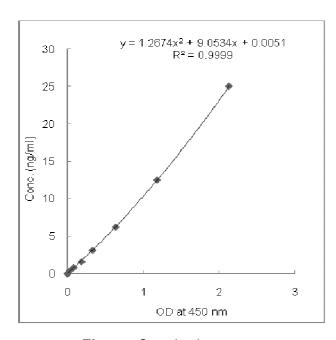
14. Measure the OD at 450 nm in an ELISA reader within 30 minutes.

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#### 11. CALCULATIONS

- Average the duplicate readings for each standard, QC and sample and subtract the average blank value (obtained with the 0 ng/ml point).
- Generate the standard curve by plotting the average absorbance obtained for each standard concentration on the horizontal (X) axis vs. the corresponding ACE2 concentration (ng/ml) on the vertical (Y) axis.
- Calculate the ACE2 concentrations of samples by interpolation of the regression curve formula as shown above in a form of a quadratic equation.
- If the test samples were diluted, multiply the interpolated values by the dilution factor to calculate the concentration of human ACE2 in the samples.

The following data are obtained using the different concentrations of standard as described in this protocol:



Standard hACE2 (ng/ml)	Optical Density (mean)		
25	2.128		
12.5	1.180		
6.25	0.635		
3.125	0.330		
1.563	0.185		
0.781	0.080		
0.391	0.032		
0	0		

Figure: Standard curve

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#### 12. PERFORMANCE CHARACTERISTICS

Typical analytical data of BioVendor human Angiotensin-Converting Enzyme 2 (ACE2) ELISA, Clinical Range are presented in this chapter

#### • Sensitivity (Limit of detection):

The lowest level of ACE2 that can be detected by this assay is 293 pg/ml. **NOTE:** The Limit of detection was measured by adding two standard deviations to the mean value of 50 zero standard.

#### Assay range:

0.391 ng/ml - 25 ng/ml

#### Specificity:

This ELISA is specific for the measurement of natural and recombinant human ACE2. It does not cross-react with human ACE, human adiponectin, human leptin, human resistin, human Nampt, human clusterin, human RBP4, human RELM-β, human IL-23, human ANG1, human ANG2, human FABP4, human ANGPTL6, human PAI-1, human vaspin, mouse RELM-α.

#### • Precision:

Intra-assay (n =11)

Six samples of known concentrations of human ACE2 were assayed in replicates 11 times to test precision within an assay.

Sample	Mean (ng/ml)	SD (ng/ml)	CV (%)	
1	1 2.785 2 4.120		6.079	
2			6.164	
3	12.680	1.257	9.914	
4	<b>4</b> 15.034		5.365	
5	24.705	1.319	5.338	
6	45.697	3.724	8.149	

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#### Inter-assay (n = 11)

Six samples of known concentrations of human ACE2 were assayed in 11 separate assays to test precision between assays.

Sample	Mean (ng/ml)	SD (ng/ml)	CV (%)
1	4.105	0.445	10.829
2	8.726	0.617	7.069
3	11.280	1.121	9.937
4	16.913	1.045	6.179
5	22.861	1.236	5.401
6	49.219	3.144	6.388

## Spinking Recovery:

When samples (urine) are spiked with known concentrations of human ACE2, the recovery averages 95% (range from 81% to 113%).

Sample	Average recovery (%)	Range (%)
1	90.8	81-97
2	93.4	84-109
3	95.1	84-111
4	95.4	89-105
5	100.7	90-113

# • Linearity

Different human urine samples containing ACE2 were diluted several fold (1 to 1/4) and the measured recoveries ranged from 89% to 107%.

Sample	Dilution	Observed (ng/ml)	Expected (ng/ml)	% of Expected
1	1:1	5.928	5.928	100
	1:2	3.027	2.964	102.1
	1:4	1.403	1.482	94.7
2	1:1	8.868	8.868	100
	1:2	4.724	4.434	106.5
	1 : 4	2.116	2.217	95.4
3	1:1	19.314	19.314	100
	1:2	10.014	9.657	103.7
	1:4	4.315	4.829	89.4

#### Expected values

ACE2 levels range in urine from 1 to > 10 ng/ml (from healthy donors).

# 13. TROUBLESHOOTING

PROBLEM	POSSIBLE CAUSES	SOLUTIONS			
	Omission of key reagent	Check that all reagents have been added in the correct order.			
	Washes too stringent	Use an automated plate washer if possible.			
No signal or weak signal	Incubation times inadequate	Incubation times should be followed as indicated in the manual.			
	Plate reader settings not optimal	Verify the wavelength and filter setting in the plate reader.			
	Incorrect assay temperature	Use recommended incubation temperature. Bring substrates to room temperature before use.			
High bookground	Concentration of detector too high	Use recommended dilution factor.			
High background	Inadequate washing	Ensure all wells are filling wash buffer and are aspirated completely.			
Poor standard	Wells not completely aspirated	Completely aspirate wells between steps.			
curve	Reagents poorly mixed	Be sure that reagents are thoroughly mixed.			
Unexpected	Omission of reagents	Be sure that reagents were prepared correctly and added in the correct order.			
results	Dilution error	Check pipetting technique and double-check calculations.			

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# 14. REFERENCES

#### References to Angiotensin-Converting Enzyme 2 (ACE2):

- 1. A human homolog of angiotensin-converting enzyme: cloning and functional expression as a captopril-insensitive carboxypeptidase: S.R. Tipnis, et al.; J. Biol. Chem. 275, 33238 (2000)
- 2. Angiotensin-converting enzyme 2-a new cardiac regulator: M. Boehm, et al.; New Eng. J. Med. 347, 1795 (2002)
- 3. Angiotensin-converting enzyme 2 is an essential regulator of heart function: M.A. Crackower, et al.; Nature 417, 822 (2002)
- 4. Angiotensin-converting enzyme 2 is a functional receptor for the SARS coronavirus: W. Li, et al.; Nature 426, 450 (2003)
- 5. Angiotensin-converting enzyme 2 protects from severe acute lung failure: Y. Imai, et al.; Nature 436, 112 (2005)

#### References to this product:

- 1. M. Kawajiri, et al.; Mult. Scler. 15, 262 (2009)
- 2. T. Matsushita, et al.; J. Neurol. Sci. 295, 41 (2010)
- 3. S. Mizuiri, et al.; Nephrol. (Carlton) (Epub ahead of print) (2011)

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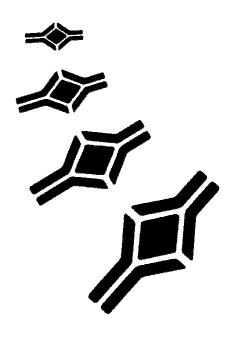
# **Assay Procedure Summary**

Prepare reagents, samples and Standards as instructed. Add 100 µl of Standards, QC and samples to each well. Incubate for 1 hour at 37°C. Aspirate and wash 3 times. Add 100 µl of Detection Antibody to each well. Incubate for 1 hour at 37°C. Aspirate and wash 3 times. Add 100 µl of diluted Detector to each well. Incubate for 1 hour at 37°C. Aspirate and wash 5 times. Add 100 µl of TMB Substrate Solution to each well. Incubate for 30 mins at RT°C in the dark. Add 100 µl of Stop Solution to each well. Read at 450 nm within 30 mins.

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