

## Preparation of samples for scanning electron microscopy (SEM)

BIOMIMESYS® Hydro scaffold is compatible with scanning electron microscopy (SEM) analyses to investigate fine structural features of the hydro scaffold and cells grown inside. Two protocols can be carried out to have an optimal conservation of the hydro scaffold (part 1) or of the cells (part 2).

### 1. Protocol for optimal conservation of the hydro scaffold structure (may alter the spheroids and cell structure)

1. Place the hydro scaffold using fine forceps in a microtube containing a cold solution of glutaraldehyde 2.5%.
2. Fixing 2 hours at 4°C
3. 3 washes with distilled or ultraclean water
4. Close the microtube containing the hydro scaffold and the distilled/ultraclean water and pierce the cap with a needle (to avoid overpressure into the tube when immersed in liquid nitrogen)
5. Then immerse the sample in the liquid nitrogen
6. Place quickly the sample in a lyophilizer and lyophilized (from overnight to 24 hours)
7. Then place the sample on a special rack for observation by scanning electron microscopy (carbon tape) and metallize prior to observation

### 2. Protocol for optimal conservation of the cell aggregates and cells structure (sample preparation alters the structure of the hydro scaffold)

1. Handle the hydro scaffold with fine forceps and place it in a microtube containing cold 2.5% glutaraldehyde (stored at 4°C)
2. Fixing 2 hours at 4°C
3. 2 washes with distilled or ultraclean water
4. Then dehydrate gradually the sample by successive baths in increasing concentrations of ethanol, at room temperature:
  - a. 30% ethanol, 30 minutes
  - b. 50% ethanol, 30 minutes 2/2

c. 70% ethanol, 30 minutes

d. 80% Ethanol 30 minutes

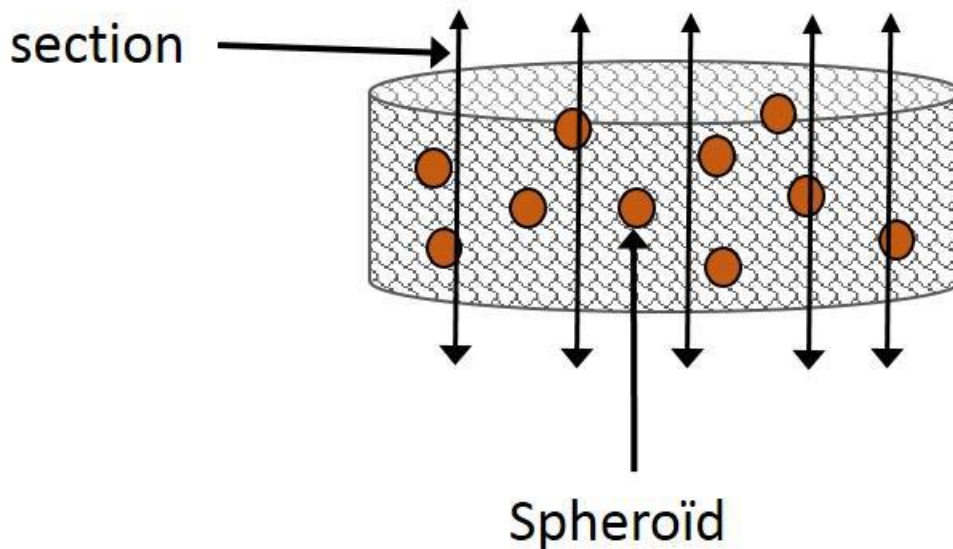
e. 90% Ethanol 30 minutes

f. 95% Ethanol, 30 minutes

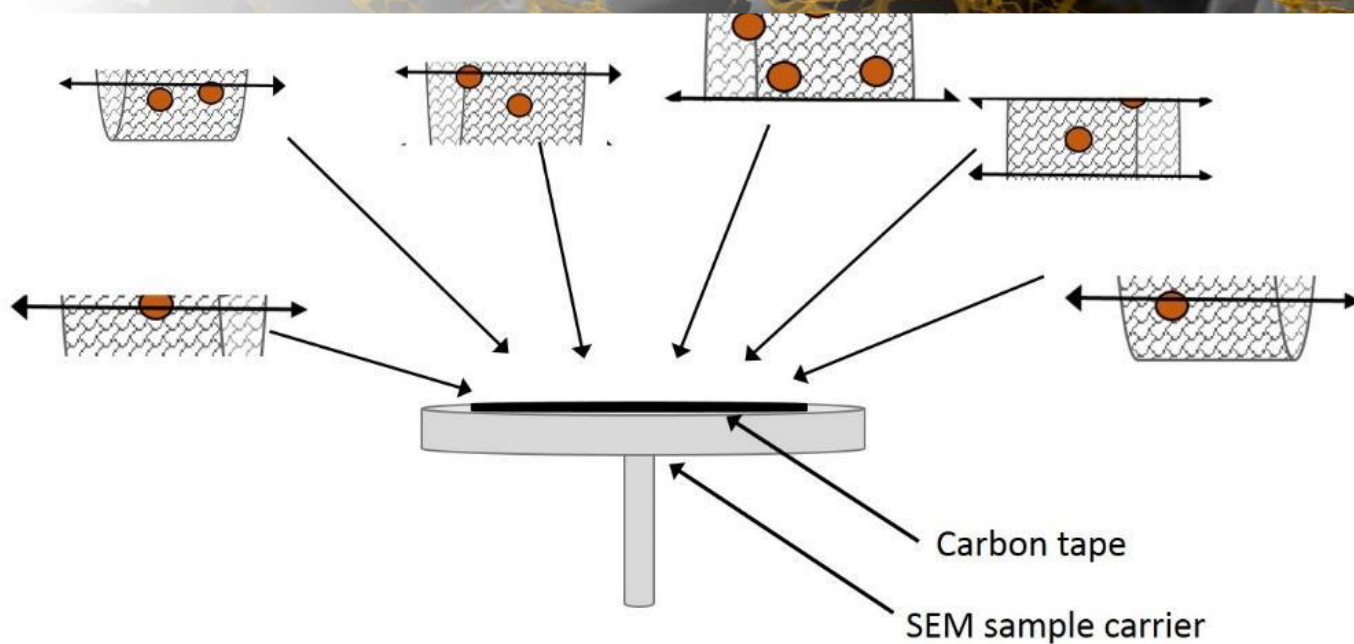
g. Absolute ethanol, 3x30 minutes

5. Place the sample under vertical flow hood overnight to achieve ethanol evaporation

6. As a result of the dehydration process, the hydrosccaffold is shrunk, as well as most cellular aggregates who had grown inside the hydrosccaffold. The SEM observation will likely require to cut the hydrosccaffold, as shown on the scheme below: Protocol for optimal conservation of the cell aggregates and cells structure (sample preparation alters the structure of the hydrosccaffold)



7. Then put down the sample pieces on the carbon tape and on the SEM sample carrier, as described below:



8. Then proceed to the metallization

9. Make your sample observations

## Contact Information

HCS Pharma

[hello@biomimesys.com](mailto:hello@biomimesys.com)

[www.biomimesys.com](http://www.biomimesys.com)