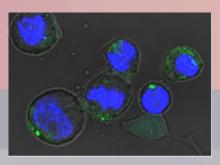
tailored transfection reagent: LeuFect-A

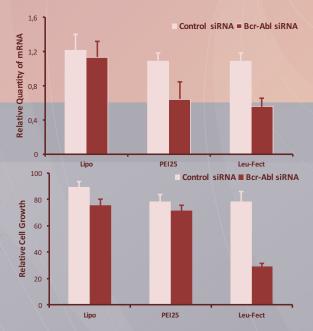
PRODUCT NUMBERS: 30-10 and 30-20	SIZE: 0.75 and 1.5 mL	CONCENTRATION: 1 mg/mL	STORAGE: -20 °C
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Product Description

LeuFect-A is a highly effective transfection reagent for attachment independent (suspension-growing) cells. **LeuFect-A** is a synthetic amphiphilic polymer that is specifically tailored for siRNA delivery. It is capable of undergoing multivalent interactions with siRNA and encapsulating co-incubated siRNA molecules into ~100 nm particles with a net positive charge. The complexation between the siRNA and **LeuFect-A** occurs in aqueous buffers, obviating the need for organic solvents during preparation. **LeuFect-A** is a non-integrating carrier of siRNA, so that the genetic make-up of host cells is not altered after treatment with the transfection reagent. **LeuFect-A** has been tested and found effective in different types of attachment-independent cells, but users are advised to test the efficacy of the reagent in their particular cell type in order to choose the right formulation for long-term use. As with all transfection reagents, formulation of **LeuFect-A** with siRNA may need to be optimized for specific cell types.



Transfecting K562 cells with siRNA using LeuFect-A and other commercial reagents. (Top, left) Intracellular delivery of fluorescently-labelled siRNA with LeuFect-A (blue: cell nucleus, green: siRNA). (Top, right) BCR-ABL mRNA analysis for cells treated with control and BCR-ABL siRNA 2 days after transfection. (Right): Growth of K562 cells after control (scrambled) and BCR-ABL siRNA delivery by LeuFect-A 3 days after transfection.



| Benefits of LeuFect-A

- High transfection efficiency in the presence of serum-containing medium.
- Simple protocol. No need to change tissue culture medium during transfection
- Minimal toxicity compared to other commercial transfection reagents, leading to better silencing.
- Non-integrating transfection reagent eliminates adverse effects due to host genome integration.

| Transfection Protocol

The following procedure is recommended for preparation of siRNA nanoparticles with **LeuFect-A** and subsequent transfection of suspension-growing cells. Please ensure all reagents are at the room temperature for the procedures.

• Recommended cell suspension is 100 to 150 x 10³ cells/mL. Cell suspension can be prepared at the desired concentration before or during the incubation of complexes.



- Recommended amounts of siRNA and LeuFect-A reagent are shown in the Table below for different multiwell plates. The final recommended concentrations of siRNA and LeuFect-A are 0.84 μg/mL and 10 μg/mL, respectively, corresponding to siRNA:LeuFect-A ratio of 1:12 and siRNA concentration of 60 nM. We recommend the siRNA and LeuFect-A concentrations to be optimized for individual cell types. Suggested ranges for optimization are 0.6 to 1.2 μg/mL for the siRNA and 6 to 15 μg/mL for the LeuFect-A. The amounts shown below are for a single well, assuming 10 μM siRNA and 1 mg/mL LeuFect-A stock solutions. Once the plate format is selected, complex volumes should be adjusted based on number of replicates.
- We recommend preparation of 10% excess volume to account for any possible loss due to pipetting.
- RPMI-1640 (or equivalent) medium without antibiotics or serum is recommended for complex preparation.

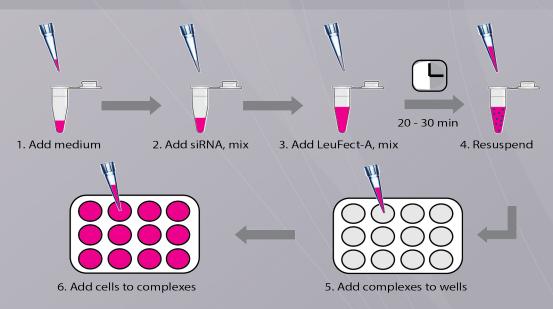
Plate Format	Medium (μL)	siRNA (μL)*	LeuFect-A (μL)*	Cell Suspension (μL)
96-well	50	0.9	1.5	100
48-well	100	2.4	4	300
24-well	200	4.8	8.1	600
6-well	500	12	20.2	1500

Recommended Volumes. * Assuming 0.14 μg/μL (10 μM) siRNA and 1 μg/μL LeuFect-A solutions.

| Procedure

- 1. Add the desired volume of medium to 1.5 mL plastic (microcentrifuge) tubes.
- 2. Add appropriate volume of siRNA solution to the medium in tubes and vortex gently for 3 sec.
- 3. Add undiluted **LeuFect-A** solution to the siRNA solution in medium. Vortex gently for 3 sec and incubate for 20-30 min at room temperature.
- 4. Re-suspend the siRNA complexes in solution using a pipette at the end of incubation.
- 5. Add complexes to the empty wells and ensure even distribution manually shake plates gently if necessary.
- 6. Carefully add the cell suspension on top of the complexes. Gently tap plate for mixing.
- 7. Incubate the plate under conditions suitable for cell culture and sample cells at desired times for analysis.

| Graphical Procedure



| References

- Gul-Uludag, et al. Leukemia Research (2014) 38: 1299-1308.
- Landry, et al. J. Controlled Release (2016) 224:8-21.
- Valencia-Serna et al. J. Controlled Release (2013) 172: 495-503.

